BIOLUMINOR

FITC-d-Lys

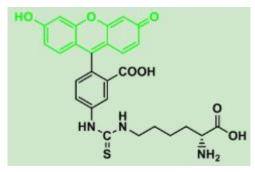
0.1 mM stock solution in anhydrous DMSO	•≤-20°C • Desiccate • Protect from light	
		• Desiccate

Table 1 Contents and storage

Introduction

By the use of the peptidoglycan biosynthetic machinery that metabolic incorporating various non-natural d-amino acids into the peptidoglycan of diverse bacteria. A chemical biology approach that enable rapid and covalent incorporation and detection of a fluorescently derivatized peptidoglycan component during cell wall synthesis in real time, in a wide range of live bacterial species. We employ metabolic incorporation of d-lysine conjugated fluoresceinisocyanate (FITC) into bacterial peptidoglycan for *in situ* probing live bacteria.

The probe is made of a fluoresceinisocyanate and d-Lysine (Fig 1), bacterias building its cell wall by the use of d-amino acids so as attache the fluorescein into the cell wall covalently. The probe not only can label gram-negative bacteria but also can label gram-positive bacteria in real time. It can be used for research and exploration infection and pathogenesis of microorganisms.



Chemical Formula: C₂₇H₂₅N₃O₇S Molecular Weight: 535.57

Fig 1. Chemical structures of FITC-d-Lys

Guidelines for Use

Before opening, allow the vial to warm to room temperature and then briefly centrifuge the vial in a micro centrifuge to deposit the DMSO solution at the bottom of the vial.

The concentration of probe for optimal staining will vary depending on the application. Here we suggest some initial conditions to use as a guideline. The staining conditions may need to be modified depending upon the particular bacteria type to the probe, among other factors.

1.1 In vitro staining FITC-d-Lys into E. coli or S. aureus

E. coli or *S. aureus* were respectively grown at 37 °C in LB medium until OD₆₀₀ reached 0.6. The medium was diluted to OD₆₀₀=0.3 with fresh medium containing FITC-d-Lys (Final concentration: 0.1 mM, usually adding 5 μ L of stock solution to per mL cell medium). The diluted bacteria were further incubation at 37°C until OD₆₀₀ = 1.0-1.5. The bacteria were centrifuged, washed with LB medium three times, and then resupended in sodium phosphate buffer (100 mM, pH 7.4) or cell culturing medium of interest. The cells were subjected to confocal fluorescence microscopy analysis (Fig 2).

(20 mM stock solution of FITC-d-Lys: 10 mg in 0.8 ml water or DMSO)

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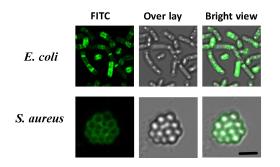


Fig 2. FITC-d-Lys labeling PG in livecells of *E. coli* and *S. aureus*. Scale bars: 5μ m.

Fluorescence spectrum

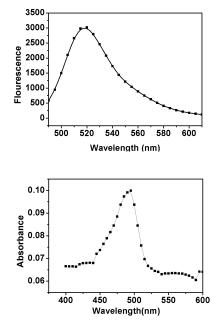


Fig 3. The Fluorescence spectra of FITC-d-Lys (1 μ M) in pH 7.4 buffer using λ_{ex} at 488 nm (A). UV-vis-NIR absorption spectra of FITC-d-Lys (1 μ M) in pH 7.4 buffer (B).

References

- 1. Angewandte Chemie International Edition, 2012, 51(50): 12519-12523.
- **2.** Nature Reviews Microbiology, 2012, 10(2): 123-136.
- 3. Analytical chemistry, 2015, 87(16): 8381-8386.